

APPLICATIONS OF SOME SMART MATERIALS BASED RELEASE DEVICES FOR HEALTH AND SAFETY PURPOSE

A K Sharma¹, B S Kaith¹

1. Dr B R Ambedkar National Institute of Technology Jalandhar – 144 011 India

ABSTRACT. In the present article, the authors have highlighted the applications of some natural backbone based smart materials for the controlled release of drugs and their antibacterial properties. Different natural backbones including Gum ghatti, Gum tragacanth and Gum salai guggal were graft copolymerized with different monomers like acrylamide, acrylic acid, methacrylic acid, aniline etc. Poly(methacrylic acid-aniline) conducting IPN was found to show non-Fickian diffusion mechanism at pH 2.2 and pH 7.0 for the controlled release of amoxicillin trihydrate. Fickian mechanism operated for the conducting hydrogel at pH 9.2. Gum ghatti-g-poly(acrylic acid-aniline) was evaluated for the release of amoxicillin trihydrate and paracetamol. The hydrogel exhibited Fickian diffusion mechanism for both the drugs at pH 2.4, pH 7.0 and pH 9.2 and can be applied for colon-specific drug delivery. Gum Salai guggal based smart material Sg-cl-polyAAM-IA-Ag⁰ was found to possess antibacterial activity against both gram (+ve) and gram (-ve) bacteria. All the synthesized smart materials were found to be effective for targeted drug delivery and antibacterial applications. Thus, the synthesized smart materials can be of great importance in health and safety sector.

Keywords: Smart materials, Antibacterial activity, Electrostatic interactions

Mr A K Sharma is a Research Scholar at Dr B R Ambedkar National Institute of Technology Jalandhar, India. His research interests are smart polymeric nanocomposites and sustained drug release devices.

Dr B S Kaith is a Professor of Chemistry at Dr B R Ambedkar National Institute of Technology Jalandhar, India. His research interests are superabsorbents, biodegradable green nanocomposites and smart nano-materials.

INTRODUCTION

Recently, smart materials based release device have become most popular for the targeted drug delivery applications as these overcome the limitations of usual administration methods [1-2]. Functional cavities of the smart materials are capable of holding the drug molecules through different type of interactions including dipole-dipole interactions, electrostatic interactions and hydrogen bonding. The interactive forces make them suitable for carrying the drug molecules to the targeted site [3]. Due to the stimuli responsive swelling behavior of smart materials, the drug molecules can be loaded over the functional cavities and can be released at the desired target site under the impact of various stimulus like pH, temperature and electric pulse [4].

The 3-D network structure of the hydrogel can be macro-porous, microporous and nanoporous depending upon the pore size of the matrix. The pore size of macro-porous hydrogels ranges from 0.1 to 1 μ m. The drug release mechanism of these hydrogels depends upon drug diffusion coefficient, tortuosity and porosity of the hydrogel matrix [5-7]. Microporous hydrogels are having small pores (100-1000 Å) and the drug releases with molecular diffusion and convection mechanism. Nanoporous hydrogels have mesh-like structure and have small pores of dimension 10-100 Å and the drug releases only via diffusion mechanism [8,9].

For drug delivery applications, hydrogel-drug complex can be formed by using a large number of crosslinking strategies including chemical-crosslinking method and UV photo polymerization technique [10]. Prior to implantation, it is necessary to remove all toxic reagents which may otherwise cause toxic impact on the targeted site. The defined dimensionality of bulk hydrogels limits their use for implantation. This problem can be solved by designing the performed hydrogel into micro or nanoparticles. Sometimes, the hydrogels can be formed *in vivo* after considering the risks of UV exposure initiation and crosslinking chemicals [11].

In this review, we highlighted the recent developments addressing the clinically relevant issues related with the use of hydrogels for drug delivery applications.

BIOMEDICAL IMPORTANCE OF SMART MATERIALS

Smart materials are biologically important materials which are designed to provide controlled drug delivery into the system of targeted site. Using smart materials as delivery vehicle, the drugs can be administered inside the body through parenteral, rectal, ocular, dermal, vaginal and nasal routes [12,13].

TYPES OF STIMULI RESPONSIVE SMART MATERIALS

pH responsive smart materials

Such type of materials responds to the change in pH of the external environment. These materials has some kind of ionisable functional groups which respond to the change in pH of external biological environment causing change in swelling/deswelling properties of the smart material.

Polyacrylic acid (PAA), polyacrylamide (PAAm), polymethylmethacrylate (PMMA) and polyethyleneglycol are the examples of some pH responsive smart materials [14]. pH sensitive smart materials are used for the delivery of insulin and drugs. The anionic copolymer of polyhydroxyethyl methyl acrylate (PHEMA) and polymethylmethacrylate showed higher swelling in basic medium and shrinkage of crosslinked matrix in acidic medium. pH and ionic strengths are the two major factors which determine the kinetics of swelling/deswelling process [15].

Temperature responsive smart materials

Temperature responsive smart materials show temperature variability of network structure in order to modulate the drug release. Most of the temperature responsive smart materials are composed of hydrophobic polymeric chains. Upon Heating, the gel matrix of negative thermo-sensitive hydrogel contract whereas positive thermos-sensitive hydrogel expands [16]. Biodegradable and biocompatible temperature responsive smart materials are used for the delivery of anticancer, antidiabetic and antibiotic drugs.

Glucose responsive smart materials

Glucose sensitive smart materials sense the presence of sugar molecules and accordingly respond for swelling / deswelling [17]. These glucose sensitive smart materials are used for implantable sensing applications to detect the sugar level inside the body and they are used for insulin delivery. Marek *et. al.*, has derived Poly(diethylaminoethyl methacrylate) Hydrogel Systems for the release of insulin and this therapeutic system controls the insulin release by change in pH or glucose concentration [18]. Some important stimuli responsive hydrogels for the release of therapeutically important drug are described in Table 1.

Wound healing properties

Smart materials are usually tissue sensitive, therefore instead of ordinary lotions, they have been designed for better patient compliance. Due to the moisturising properties of the smart materials, dryness and scaling is not happening with the drug delivery of such type of systems [30], [31]. Hydrogels possess wound healing properties due to their biocompatibility with the natural tissues. Gum *Salai guggal* based semi-IPN nanocomposites possessed antibacterial activity due to the presence of nanosilver present inside the crosslinked semi-IPN matrix. The nanosilver was found to possess an average particle size of 5 ± 1 nm. Gum *Salai guggal* based nanocomposites can be used as a scaffolds against bacterial infection caused by gram (+ve) and gram (-ve) bacterial strains [32]. Ggum-poly(Itaconic Acid) based superabsorbents possess wound healing properties due to their antibacterial activity against gram positive bacteria *S. aureus* [33]. Gum xanthan is a high molecular weight polysaccharide which is obtained from the fermentation of glucose, sucrose or similar complex substrate obtained from sugarcane, corn or similar complex. The crosslinking of acrylic acid onto the gum xanthan results into crosslinked hydrogel Gx-cl-poly(AA)-MW. The synthesised hydrogel possessed antibacterial activity against *Bacillus*

subtilis and *Salmonella enteritidis* [34]. The inhibited bacterial zone of test samples against different bacterial strains is summarized in Table 2.

Table 1: Some therapeutically important smart materials for the release of drugs

Stimulus	Smart Materials	Therapeutic moieties	Reference(s)
pH	Tri polymer of N-vinyl 2- pyrrolidone methacrylamide and itaconic acid.	Insulin	[19]
pH	Copolymer of polymethacrylic acid and polyethylene glycol	Calcitonin	[20], [21]
pH	Copolymer of cationic guar gum and acrylic acid monomer	Ketoprofen	[22]
pH	Polyethyleneglycol.	Camptothecin	[23]
pH	poly(ethylene glycol) monomethyl ether monomethacrylate (PEGME)	N-(4-hydroxyphenyl)-2-(4-methoxyphenyl) acetamide	[24]
Temperatue	Co-polymer of poly-PNIPA and poly-PNIPA-Co-AA	5-Flurouracil	[25]
Temperatue	NIPAAm-Co-AAm	Vaginal Microbicide	[26]
Temperatue	Copolymer of gelatin and PVA	Adrenochrome	[27]
Temperatue	NIPAAm-Co.AAm	Insulin	[28]
Glucose	poly(hydroxyethyl methacrylate) (HEMA)	protein	[29]

Target oriented drug delivery devices

The selective delivery of drug to the desired target site can be done through multifunctional smart polymeric materials. Smart materials based drug delivery is more target specific compared to ordinary drug delivery systems. Gamma radiation induced synthesis of semi-IPN matrices based on agar and gelatin was found to be effective for colon and intestine specific drug delivery of diethyl carbamazine citrate and amoxicillin. The drug release was found to be non-Fickian in pH 7.0, 9.2 and Fickian in pH 2.2 for carbamazine citrate drug diffusion. The drug amoxicillin showed case II diffusion in pH 9.2. Non-Fickian type of diffusion mechanism was followed for the release of drug amoxicillin in pH 9.2 and 7.0 [35]. An interpenetrating network of Gum ghatti-graft-poly(methacrylic acid-aniline) can also be used as controlled release device for the delivery of amoxicillin trihydrate. The release of drug was found to be higher in basic medium

i.e. pH 9.2 as compare to pH 7.0 and 2.2. The device was target oriented for colon specific drug delivery of amoxicillin in the lower gastrointestinal tract [36]. Gum tragacanth and acrylic acid based hydrogels Gt-cl-poly(AA) were found to be suitable for the delivery of antiulcerative drug pantoprazole sodium.

Table 2: Zone of inhibition of synthesized samples against different bacterial strains

Sample code	Bacteria	Zone of inhibition (mm)	Reference(s)
Sg-cl-polyAAm-IA-Ag ⁰	<i>P. aeruginosa</i>	25 ± 1	[32]
Sg-cl-polyAAm-IA-Ag ⁰	<i>B. cereus</i>	22 ± 1	[32]
Sg-cl-polyAAm-IA-Ag ⁰	<i>S. aureus</i>	13 ± 0.5	[32]
Sg-cl-polyAAm-IA-Ag ⁰	<i>E. coli</i>	14 ± 1	[32]
Ggum-cl-poly(IA)	<i>S. aureus</i>	22.5	[33]
Ggum-cl-poly(IA-ipn-ANI)-neutral	<i>S. aureus</i>	18.2	[33]
Ggum-cl-poly(IA-ipn-ANI)-acidic	<i>S. aureus</i>	16	[33]
Gx-cl-poly(AA)-MW	<i>Bacillus subtilis</i>	14.17 ± 0.12	[34]
Gx-cl-poly(AA)-MW	<i>Salmonella enteritis</i>	13.66 ± 0.54	[34]

The pH sensitive hydrogel Gt-cl-poly(AA) was found to be suitable for colon-specific drug delivery. Since, the final drug release was achieved after 30 hrs, therefore the device was found to be effective for prolonged drug delivery [37]. Gum tragacanth based binary graft copolymer poly[(acrylic acid)-co-acrylamide] was found to be suitable for the controlled release of antihypertensive drug losartan potassium. This device showed final release of the drug after 34 hrs indicating its applicability for the prolonged release of the drug losartan potassium [38].

CONCLUSION

Smart materials based release devices derived from natural backbones have sufficient potential against different gram (+ve) and gram (-ve) bacterial strains and can be used as scaffolds for the treatment of infected wounds. These device can also be used for the controlled and sustained release of different drugs and thus can be used for health and safety purpose.

REFERENCES

- [1] H. Priya James, R. John, A. Alex, and K. R. Anoop, "Smart polymers for the controlled delivery of drugs – a concise overview," *Acta Pharm. Sin. B*, vol. 4, no. 2, pp. 120–127, 2014.
- [2] G. G. Genchi, A. Marino, C. Tapeinos, and G. Ciofani, "Smart Materials Meet Multifunctional Biomedical Devices: Current and Prospective Implications for Nanomedicine," *Front. Bioeng. Biotechnol.*, vol. 5, 2017.
- [3] K. V. Ramana Reddy, M. V. Nagabhushanam, and P. S. Chowdary, "Role of polymers and their role in functional aspects of swellable mucoadhesive hydrogel beads - A review," *Indo Am. J. Pharm. Res.*, vol. 6, no. 01, pp. 4046–4054, 2016.
- [4] N. A. Jalili, M. K. Jaiswal, C. W. Peak, L. M. Cross, and A. K. Gaharwar, "Injectable nanoengineered stimuli-responsive hydrogels for on-demand and localized therapeutic delivery," *Nanoscale*, vol. 9, no. 40, pp. 15379–15389, 2017.
- [5] A. Serrano-Aroca, J. L. Gomez Ribelles, M. M. Pradas, A. Vidaurre, A. S. Gimeno, "Characterization of macroporous polymethyl methacrylate coated with plasma polymerized poly 2- hydroxyethyl acrylate," *Eur. Polym. J.*, vol. 43, pp. 4552–4564, 2007.
- [6] J. A. Rowley, G. Madlambayan, and D. J. Mooney, "Alginate hydrogels as synthetic extracellular matrix materials," *Biomaterials*, vol. 20, no. 1, pp. 45–53, 1999.
- [7] Q. Liu, E. L. Hedberg, Z. Liu, R. Bahulekar, R. K. Meszlenyi, and A. G. Mikos, "Preparation of macroporous poly(2-hydroxyethyl methacrylate) hydrogels by enhanced phase separation," *Biomaterials*, vol. 21, no. 21, pp. 2163–2169, 2000.
- [8] M. E. McNeill and N. B. Graham, "Properties controlling the diffusion and release of water-soluble solutes from poly(ethylene oxide) hydrogels 2. Dispersion in an initially dry slab," *J. Biomater. Sci. Polym. Ed.*, vol. 5, no. 1–2, pp. 111–130, 1994.
- [9] M. S. Jhon and J. D. Andrade, "Water and hydrogels," *J. Biomed. Mater. Res.*, vol. 7, no. 6, pp. 509–522, 1973.
- [10] T. R. Hoare and D. S. Kohane, "Hydrogels in drug delivery: Progress and challenges," *Polymer (Guildf.)*, vol. 49, no. 8, pp. 1993–2007, 2008.
- [11] M. M. Talukdar, I. Vinckier, P. Moldenaers, and R. Kinget, "Rheological characterization of xanthan gum and hydroxypropylmethyl cellulose with respect to controlled-release drug delivery," *J. Pharm. Sci.*, vol. 85, no. 5, pp. 537–540, 1996.
- [12] M. R. Kim and T. G. Park, "Temperature-responsive and degradable hyaluronic acid / Pluronic composite hydrogels for controlled release of human growth hormone," *J. Control. Release*, vol. 80, pp. 69–77, 2002.

- [13] T. Matsuda, "Device-directed therapeutic drug delivery systems," *J. Control. Release*, vol. 78, no. 1–3, pp. 125–131, 2002.
- [14] L. Liu, W. D. Yao, Y. F. Rao, X. Y. Lu, and J. Q. Gao, "pH-responsive carriers for oral drug delivery: Challenges and opportunities of current platforms," *Drug Deliv.*, vol. 24, no. 1, pp. 569–581, 2017.
- [15] L. Brannon-Peppas and N. A. Peppas, "Dynamic and equilibrium swelling behaviour of pH-sensitive hydrogels containing 2-hydroxyethyl methacrylate¹," *Biomaterials*, vol. 11, no. 9, pp. 635–644, 1990.
- [16] K. S. Soppimath, T. M. Aminabhavi, A. M. Dave, S. G. Kumbar, and W. E. Rudzinski, "Stimulus-responsive 'smart' hydrogels as novel drug delivery systems," *Drug Dev. Ind. Pharm.*, vol. 28, no. 8, pp. 957–974, 2002.
- [17] D. T. Eddington and D. J. Beebe, "Flow control with hydrogels," *Adv. Drug Deliv. Rev.*, vol. 56, no. 2, pp. 199–210, 2004.
- [18] S. R. Marek and N. A. Peppas, "Insulin release dynamics from poly(diethylaminoethyl methacrylate) hydrogel systems," *AIChE J.*, vol. 59, no. 10, pp. 3578–3585, 2013.
- [19] N. Sood, A. Bhardwaj, S. Mehta, and A. Mehta, "Stimuli-responsive hydrogels in drug delivery and tissue engineering," *Drug Deliv.*, vol. 23, no. 3, pp. 758–780, 2016.
- [20] A. Serres, M. Baudys, and S. W. Kim, "Temperature and pH sensitive polymers for Human Calcitonin Delivery," *Pharm. Res.*, vol. 13, no. 2, pp. 196–198, 1996.
- [21] M. Torres-lugo and N. A. Peppas, "Molecular Design and in Vitro Studies of Novel pH-Sensitive Hydrogels for the Oral Delivery of Calcitonin," pp. 6646–6651, 1999.
- [22] K. Takayama and T. Nagai, "Simultaneous optimization for several characteristics concerning percutaneous absorption and skin damage of ketoprofen hydrogels containing d-limonene," *Int. J. Pharm.*, vol. 74, no. 2–3, pp. 115–126, 1991.
- [23] A. Lalloo, P. Chao, P. Hu, S. Stein, and P. J. Sinko, "Pharmacokinetic and pharmacodynamic evaluation of a novel in situ forming poly(ethylene glycol)-based hydrogel for the controlled delivery of the camptothecins.," *J. Control. Release*, vol. 112, no. 3, pp. 333–42, 2006.
- [24] T. Bartil, M. Bounekhel, C. Cedric, and R. Jeerome, "Swelling behavior and release properties of pH-sensitive hydrogels based on methacrylic derivatives," *Acta Pharm.*, vol. 57, no. 3, pp. 301–314, 2007.
- [25] H. Chen, Y. Gu, Y. Hub, and Z. Qian, "Characterization of {pH}- and temperature-sensitive hydrogel nanoparticles for controlled drug release," *PDA J. Pharm. Sci. Technol.*, vol. 61, no. 4, pp. 303–313, 2007.

- [26] K. M. Gupta, S. R. Barnes, R. A. Tangaro, M. C. Roberts, D. H. Owen, D. F. Katz, and P. F. Kiser, "Temperature and pH sensitive hydrogels: An approach towards smart semen-triggered vaginal microbicides," *J. Pharm. Sci.*, vol. 96, no. 3, pp. 670–681, 2007.
- [27] D. Mukherjee and A. K. Banthia, "Preparation of adrenochrome hydrogel patch, gel, ointment, and the comparison of their blood coagulating and wound healing capability," *Mater. Manuf. Process.*, vol. 21, no. 3, pp. 297–301, 2006.
- [28] M. Bikram, A. M. Gobin, R. E. Whitmire, and J. L. West, "Temperature-sensitive hydrogels with SiO₂-Au nanoshells for controlled drug delivery," *J. Control. Release*, vol. 123, no. 3, pp. 219–227, 2007.
- [29] A. A. Obaidat and K. Park, "Characterization of protein release through glucose-sensitive hydrogel membranes," *Biomaterials*, vol. 18, no. 11, pp. 801–806, 1997.
- [30] N. Trookman, R. Rizer, T. J. Stephens, and R. Trancik, "Atopic dermatitis: Advantages of a novel hydrogel vehicle," *J. Am. Acad. Dermatol.*, vol. 56, no. 2, p. AB75, 2007.
- [31] A. Wynne, M. Whitefield, A. Dixon, and S. Anderson, "An effective, cosmetically acceptable, novel hydro-gel emollient for the management of dry skin conditions," *J. Dermatolog. Treat.*, vol. 13, no. 2, pp. 61–66, 2002.
- [32] A. K. Sharma, B. S. Kaith, B. Gupta, U. Shanker and S. P. Lochab, "A facile strategy to synthesize a novel and green nanocomposite based on gum Salai guggal - Investigation of antimicrobial activity". *Mater. Chem. Phys.*, vol. 219, pp. 129-141, 2018.
- [33] R. Sharma, S. Kalia, B. S. Kaith, A. Kumar, P. Thakur, D. Pathania, and M. K. Srivastava, "Ggum-poly(Itaconic Acid) Based Superabsorbents Via Two-Step Free-Radical Aqueous Polymerization for Environmental and Antibacterial Applications," *J. Polym. Environ.*, vol. 25, no. 2, pp. 176–191, 2017.
- [34] Sukriti, B. S. Kaith, R. Jindal, M. Kumari, and M. Kaur, "Biodegradable-stimuli sensitive xanthan gum based hydrogel: Evaluation of antibacterial activity and controlled agro-chemical release," *React. Funct. Polym.*, vol. 120, pp. 1–13, 2017.
- [35] P. Mehta and B. S. Kaith, "Gamma radiative fabrication of semi interpenetrating network film: Optimization, characterization and investigation as colon, intestine specific drug release device," *Vacuum*, vol. 156, pp. 357–369, 2018.
- [36] K. Sharma, V. Kumar, B. S. Kaith, S. Som, V. Kumar, A. Pandey, S. Kalia, and H. C. Swart, "Synthesis of biodegradable Gum ghatti based poly(methacrylic acid-aniline) conducting IPN hydrogel for controlled release of amoxicillin trihydrate," *Ind. Eng. Chem. Res.*, vol. 54, no. 7, pp. 1982–1991, 2015.

- [37] Saruchi, B. S Kaith, R. Jindal and G. S. Kapur, "Synthesis of Gum tragacanth and acrylic acid based hydrogel: its evaluation for controlled release of antiulcerative drug pantoprazole sodium," *J. Chinese Adv. Mater. Soc.*, vol. 2, no. 2, 2014.
- [38] Saruchi, B. S. Kaith, R. Jindal, V. Kumar, and M. S. Bhatti, "Optimal response surface design of Gum tragacanth-based poly[(acrylic acid)-co-acrylamide] IPN hydrogel for the controlled release of the antihypertensive drug losartan potassium," *RSC Adv.*, vol. 4, no. 75, pp. 39822–39829, 2014.